

# Simulating T-wave parameters of local extracellular electrograms with a whole-heart bidomain reaction-diffusion model: Size matters!

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**Abstract**—As a measure of local repolarization time ( $T_R$ ), the instant of maximum slope ( $T_{up}$ ) of the T wave in the local unipolar electrogram is commonly used. Measurement of  $T_{up}$  can be difficult, especially in positive T waves. These difficulties have led some researchers to propose the instant of maximum downslope ( $T_{down}$ ) as a marker of  $T_R$  when the T wave is positive. To improve understanding of T-wave parameters, we simulated electrograms with a bidomain model of the human heart. To test T-wave parameters, we compared them to  $T_R$  determined from the local membrane potential. We propose a simple model of the electrogram, which we validated by comparison to the bidomain model. With the simple model, it is straightforward to show that the sign of the T wave is almost uniquely determined by  $T_R$ . We then used the bidomain model to simulate the effects of a variety of pathologies and technical difficulties, which the simple model could not account for. Generally,  $T_{up}$  was a much better estimate for  $T_R$  than  $T_{down}$ . Regional fibrosis could attenuate local electrogram components and reduce accuracy of  $T_{up}$  as a marker for  $T_R$ . In fibrotic tissue,  $T_{down}$  was not related to  $T_R$  at all. This investigation of electrogram slopes required the simulation of extracellular potentials with about 100 times more precision than needed for simulation of visually acceptable waveforms alone. This requirement is more difficult to meet in larger models, but it was actually possible for a human-heart model with 60 million nodes. By sacrificing some spatial resolution, we kept the computational requirements within acceptable limits for multiple simulations.

## I. INTRODUCTION

It is difficult if not impossible to measure transmembrane potentials ( $V_m$ ) at many sites simultaneously in a beating heart. Extracellular electrograms are a convenient alternative. Local activation times are easily determined using the instant of steepest downstroke in a (unipolar) electrogram. The action potential duration (APD) is more difficult to measure. Wyatt et al. proposed the activation-recovery interval (ARI) as a substitute [1]. The ARI was measured from the instant of steepest downstroke to the instant of steepest upstroke ( $T_{up}$ ) of the T wave in the unipolar electrogram. Experimental and theoretical studies confirmed the validity of this method [2], [3], [4], [5].

Several authors have proposed that an exception should be made for positive T waves, using the instant of steepest downstroke ( $T_{down}$ ) of the T wave in the unipolar electrogram instead of  $T_{up}$  [6], [7]. In contrast to the original

ARI method, the method based on  $T_{down}$  lacks a theoretical foundation. Yue et al. suggested that the derivation of the original method, being based on one-dimensional (1-D) considerations, would be invalid in the 3-D heart [7].

In this study, we first show that the T wave can be understood as a weighted sum of the local  $V_m$  and a remote component. It follows that local repolarization always causes a rise in the local electrogram. We define repolarization time ( $T_R$ ) as the instant of steepest downstroke of  $V_m$ . In the absence of remote activity,  $T_{up}$  equals  $T_R$ . In order to estimate how much distortion can be expected due to remote activity, we compared  $T_R$  computed from  $V_m$  to  $T_{up}$  computed from electrograms simulated with a 3-D model of the human heart. Simulations were performed with regions of abnormally long and short APD, and with a representation of fibrotic tissue.

## II. METHODS

Propagating  $V_m$  were simulated with a monodomain reaction-diffusion model of the human heart [8]. This model has anisotropic myocardium and includes five different types of myocytes (Tab. I). In order to mimic sinus rhythm we stimulated the ventricles at the early activation sites published by Durrer et al. [9]. Ionic currents were computed with the 2004 version of the TNNP model for the human ventricular myocyte [10]. Some parameters of the ionic model were changed, and differences between the left ventricle (LV) and right ventricle (RV) were implemented as outlined in Tab. I. Modifications were made according to published data (on canine hearts) [11], [12]. The types XL and XS were used to implement abnormally long and short APD in some experiments.

Two different models were used for the computation of extracellular potentials ( $\phi_e$ ): a “realistic model” and a “simple model.”

*a) realistic model:* For maximal realism,  $\phi_e$  was computed from  $V_m$  throughout the heart by solving

$$\nabla \cdot ((G_i + G_e)\nabla\phi_e) = -\nabla \cdot (G_i\nabla V_m) \quad (1)$$

where  $G_i$  and  $G_e$  are the intracellular and extracellular conductivity tensor fields, respectively [8]. In our previous work we have done this at a spatial resolution of 0.2 mm and with error tolerance levels for  $\phi_e$  that were just small enough to produce correct electrograms. However, the temporal derivatives of the signals so computed contain too much distortion for the current study. Therefore the tolerance levels were set a factor 100 lower (see our previous work for the definition of the two tolerance levels involved [8]). At a

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TABLE I  
SELECTED PARAMETERS AND INTRINSIC CHARACTERISTICS OF THE IONIC MODEL.

	LV epi	LV M	(LV&RV) endo	RV M	RV epi	XS	XL
$G_{to}$ (nS/pF)	0.294	0.294	0.073	<b>0.504</b>	<b>0.882</b>	0.294	0.073
$G_{Ks}$ (nS/pF)	0.245	0.062	0.245	<b>0.112</b>	<b>0.490</b>	<b>0.735</b>	<b>0.010</b>
$G_{Kr}$ (nS/pF)	0.096	0.096	0.096	0.096	0.096	0.096	<b>0.020</b>
APD (ms)	272	324	275	303	244	218	443
APD20 (ms)	171	201	150	205	183	126	207
APD50 (ms)	250	300	251	282	227	197	402
APD70 (ms)	267	319	270	299	241	213	435
APD90 (ms)	278	330	281	309	251	225	449
DR	82 %	82 %	82 %	81 %	80 %	81 %	83 %

Parameter values that are different from the original TNNP model [10] are printed in bold type. APD $x$  = action potential duration at  $x$  percent repolarization (1000 ms cycle length). DR = the degree of repolarization at the end of the action potential (defined as the instant of steepest downslope). Units are nS = nanoSiemens, pF = picoFarad, ms = millisecond.

spatial resolution of 0.2 mm this led to one week computation time on 32 processors, which is too much for repeated simulations. Therefore we chose a lower spatial resolution of 0.25 mm for computation of both  $V_m$  and  $\phi_e$ . This leads to somewhat slower propagation, for which we corrected by choosing 10–15 % higher tissue conductivity values for the intracellular domain. Nominal conductivities were taken from Roth [13]. The adapted values were  $\sigma_{eT} = 0.12$ ,  $\sigma_{eL} = 0.3$ ,  $\sigma_{iT} = 0.035$  and  $\sigma_{iL} = 0.33 \text{ Sm}^{-1}$ , with subscript ‘e’ for extracellular, ‘i’ for intracellular, ‘T’ for transverse, and ‘L’ for longitudinal.

*b) simple model:* If it is assumed that the heart and intracavitary blood are uniformly isotropic and that the reference point is equally well connected with any position in the ventricles, then the unipolar electrogram at a point  $x$  is simply a scaled mirror image of the difference between the local  $V_m$  and the average  $V_m$  in the ventricular myocardium ( $V_{avg}$ ):

$$\phi_{e,\text{simple}}(x) = -\frac{\sigma_i}{\sigma_i + \sigma_e} (V_m(x) - V_{avg}) \quad (2)$$

where  $\sigma_e$  and  $\sigma_i$  represent the conductivities of the extracellular and intracellular domains, respectively. The “simple model” uses this formula, evaluating  $V_{avg}$  at 2-mm resolution in the ventricles. For the fraction  $\sigma_i/(\sigma_i + \sigma_e)$  the value 0.25 was chosen.

The reference potential for electrograms was taken from the roof of the right atrium. Simulations were performed with a normal-heart model and models containing a modified zone of 10 mm radius (3 cm<sup>3</sup> volume, 1.2 % of the myocardium) located in the LV free wall. This zone had either abnormally short or abnormally long APD, with either normal or fibrotic tissue. Diffuse fibrosis was represented macroscopically by a reduction of the intracellular volume fraction from its assumed normal value of 0.7 to 0.1. This change was represented in the model by effective tissue conductivity values that were adapted according to the modified cross-sectional surfaces of the two domains. The intracellular  $\sigma$  were multiplied by  $(0.1/0.7)^{2/3}$ , and the extracellular  $\sigma$  by  $(0.9/0.3)^{2/3}$ . The resulting values were  $\sigma_{eT} = 0.250$ ,  $\sigma_{eL} = 0.624$ ,  $\sigma_{iT} = 0.0082$  and  $\sigma_{iL} = 0.082 \text{ Sm}^{-1}$ .

T waves could be positive, negative, biphasic, or multiphasic. To allow a division into positive and negative T waves, we used the integral of the electrogram from 100 ms after local depolarization to the end of the simulation. A T wave was defined as positive when the integral was positive. Repolarization time ( $T_R$ ) was defined as the instant of steepest downstroke of  $V_m$ .  $T_R$  and  $T_{up}$  were evaluated in the interval from 100 ms after local depolarization to the end of the simulation. For positive T waves,  $T_{down}$  was evaluated in the interval from  $T_{up}$  to the end of the simulation. For negative T waves,  $T_{down}$  was not defined. Analysis was fully automatic.

### III. RESULTS

#### A. Validation of the simple model

Fig. 1, panel A, shows  $V_m$ ,  $V_{avg}$ , and the electrogram according to the “simple model” for one site in the heart. Temporal derivatives are also shown. The derivative of  $V_m$  reaches a lower value than the derivative of  $V_{avg}$ , so it dominates  $T_{up}$  of the local electrogram. However, the derivative of  $V_{avg}$  is not negligible and may influence  $T_{up}$ . In panel B, electrograms are compared that were computed with the “simple” and “realistic” models. These simulated electrograms were highly similar. In particular, both models agreed on the sign of the T wave in 92 % of the analyzed positions (panel C).

These comparisons show that the T wave is essentially determined by the local  $V_m$  and by  $V_{avg}$ . The electrogram is positive when the local  $V_m$  is lower than  $V_{avg}$ . This happens in particular for early-repolarizing cells. The electrogram remains positive as long as there are depolarized cells elsewhere in the heart. Conversely, the electrogram will be negative when local  $V_m$  is still depolarized while  $V_m$  elsewhere is repolarized.

Fig. 2 shows electrograms according to the simple model and the realistic model at a position where a thin strand of myocardium is surrounded by intracavitary blood and connective tissue. Here, the realistic model predicts a low-amplitude electrogram with a short downstroke and a T wave that is dominated by the remote component. The simple model does not account for the small muscle mass in a large

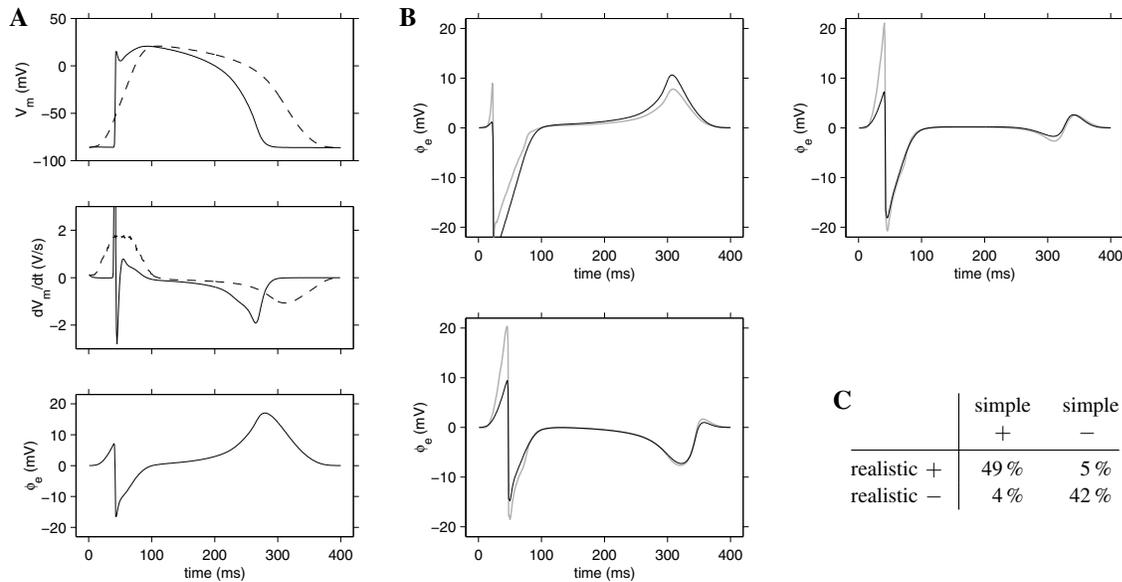


Fig. 1. Comparison between the simple and realistic models of the unipolar electrogram. **Panel A** shows how an electrogram is reconstructed according to the “simple model.” The top panel shows  $V_{avg}$  (dashed line) and local  $V_m$  (drawn line). The middle panel shows their temporal derivatives (dashed for  $dV_{avg}/dt$ ). The lower panel shows the reconstructed electrogram. In **panel B**, electrograms according to the simple model (black lines) are compared to the realistic model (gray lines). In **panel C**, T-wave sign according to the two models is compared for a sample of 1000 positions.

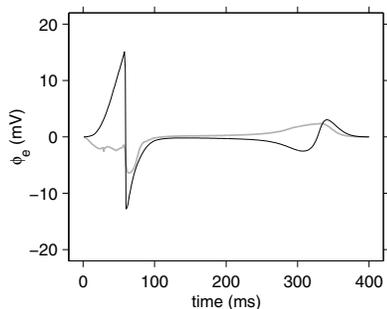


Fig. 2. Electrograms according to the simple (black) and realistic model (gray) at a position where a thin strand of myocardium is surrounded by intracavitary blood and connective tissue.

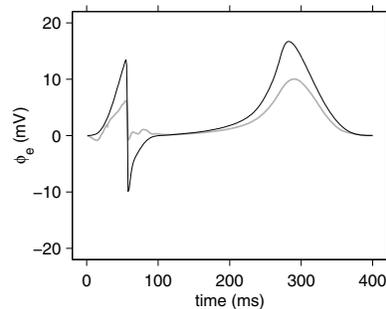


Fig. 3. Electrograms according to the simple model (black line) and the realistic model (gray line) in fibrotic tissue.

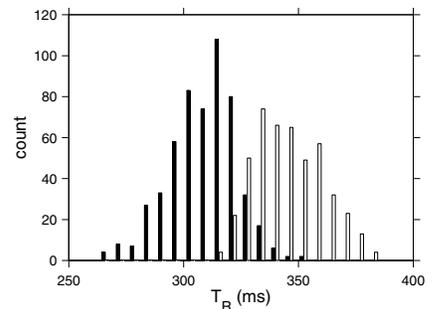


Fig. 4. Distribution of  $T_R$  for positive T waves (black bars) and for negative T waves (white bars).

extracellular space, and predicts a normal high-amplitude electrogram. According to the realistic model, the difference between  $T_{up}$  and  $T_R$  would be 33 ms, while the simple model predicts no error. A typical example of simulated electrograms in a fibrotic region is shown in Fig. 3. Here too the two models predict different R waves and downstrokes, but they agree on T-wave upstroke timing.

### B. T-wave sign in the normal heart

The following results were obtained with the “realistic model.” In the normal heart, positive T waves were found in 54 % of the analyzed positions, and were associated with a 36 ms earlier  $T_R$  than negative T waves. In Fig. 4,  $T_R$  distribution is shown separately for positive and negative T waves. Most positive T waves were associated with  $T_R$  that were lower than those of negative T waves, but some overlap between the two distributions was present.

### C. Repolarization statistics

Figure 5 shows a sample of electrograms taken from various sites in the heart, selected to show the variation in T-wave shape from entirely negative through biphasic to positive. Local repolarization times ( $T_R$ ) are indicated with dots in the electrograms. These are invariably located on the upslope of the T wave.

Tab. II shows paired comparisons of repolarization characteristics, for both positive and negative T waves. Differences were computed for individual positions, then the average and standard deviation of the difference were computed. For positive T waves,  $T_{up}$  underestimated  $T_R$  by  $0.1 \pm 2.2$  ms and  $T_{down}$  overestimated  $T_R$  by  $29.3 \pm 7.8$  ms.

The relation between  $T_R$  and electrogram-based measures was also assessed by correlation analysis and by a linear fit. For negative T-wave morphologies,  $T_{up}$  correlated very well with  $T_R$  ( $r = 0.996$ ). The slope of the regression line was 1.012. For positive T waves the correlation was 0.988, and

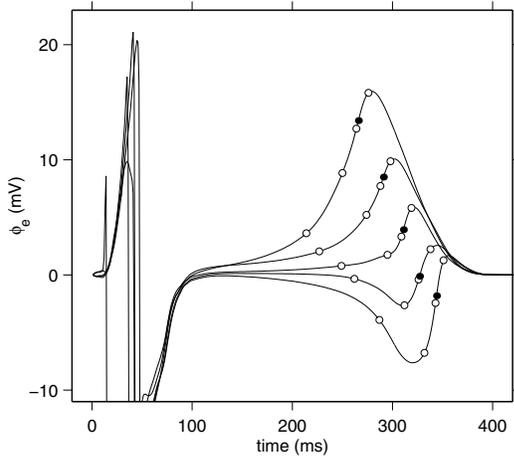


Fig. 5. Simulated electrograms from various sites in the heart. Local  $T_R$  are indicated with dots. The instants of 20, 50, 70, and 90% repolarization are indicated with open circles.

TABLE II  
COMPARISON OF REPOLARIZATION MEASURES (MS).

	positive T	negative T
whole ventricles:		
$T_{up} - T_R$	$-0.1 \pm 2.2$	$0.7 \pm 1.7$
$T_{down} - T_R$	$29.3 \pm 7.8$	undefined
fibrotic sites:		
$T_{up} - T_R$	$-2.9 \pm 9.4$	none
$T_{down} - T_R$	$54.3 \pm 16.5$	undefined

slope 0.986. The residual error of the linear fit was 2 ms for both positive and negative T waves. Correlation between  $T_{down}$  and  $T_R$  was 0.853, and associated with a slope of only 0.625, with a residual error of 6 ms.

A simulation was performed with a small area in which cells had a very short APD (type XS in Tab. I). Statistics were compared with those of the normal heart. While differences in  $T_{up}$  ( $\Delta T_{up}$ ) correlated well ( $r = 0.989$ ) with differences in  $T_R$  ( $\Delta T_R$ ), the differences in  $T_{down}$  ( $\Delta T_{down}$ ) were more weakly related ( $r = 0.802$ ).

In fibrotic tissue with short APD,  $T_{up}$  became biased by  $-3$  ms and was associated with a 9 ms s.d. error (Tab. II). Correlation between  $T_{up}$  and  $T_R$  was 0.85.  $T_{down}$  completely lost its relation to  $T_R$  and became nearly uniform throughout the fibrotic area ( $r = 0.21$ , slope 0.052).

#### IV. DISCUSSION

The recent controversy on repolarization measurement in the unipolar electrogram (UEG) demonstrates that T-wave polarity in the UEG is badly understood. We have shown that at least for T waves in healthy tissue the UEG can be understood as a scaled difference between the local  $V_m$  and the average  $V_m$  in the heart. With this simple model, T-wave polarity is easy to understand. The UEG is positive when the local  $V_m$  is more negative than the average, and negative when the local  $V_m$  is more positive. The most early-repolarizing sites are therefore characterized by positive T waves. Later sites have an initially negative T wave, due

to the decrease of the average potential caused by the earlier sites. When they repolarize, their  $V_m$  quickly becomes more negative than the average, causing a rapid change in their UEG from negative to positive. Only the latest repolarizing sites have entirely negative T waves.

Our simulations confirm that  $T_{up}$  is a good estimate for  $T_R$ . We have tested its validity in a variety of difficult conditions. Strong local dispersion of APD could reduce the accuracy of  $T_{up}$  as a measure of  $T_R$ . However, its accuracy was always better than that of  $T_{down}$ .

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#### REFERENCES

- [1] R. F. Wyatt, M. J. Burgess, A. K. Evans, R. L. Lux, J. A. Abildskov, and T. Tsutsumi, "Estimation of ventricular transmbrane action potential durations and repolarization times from unipolar electrograms," *Am. J. Cardiol.*, vol. 47 (Part II), p. 488, 1981, (abstract).
- [2] C. K. Millar, F. A. Kralios, and R. L. Lux, "Correlation between refractory periods and activation-recovery intervals from electrograms: effects of rate and adrenergic interventions," *Circulation*, vol. 72, pp. 1372–1379, 1985.
- [3] C. W. Haws and R. L. Lux, "Correlation between in vivo transmbrane action potential durations and activation-recovery intervals from electrograms. Effects of interventions that alter repolarization time," *Circulation*, vol. 81, pp. 281–288, 1990.
- [4] R. Coronel, J. M. T. de Bakker, F. J. G. Wilms-Schopman, T. Opthof, A. C. Linnenbank, C. N. Belterman, and M. J. Janse, "Monophasic action potentials and activation recovery intervals as measures of ventricular action potential duration: Experimental evidence to resolve some controversies," *Heart Rhythm*, vol. 3, pp. 1043–1050, 2006.
- [5] B. M. Steinhaus, "Estimating cardiac transmbrane activation and recovery times from unipolar and bipolar extracellular electrograms: A simulation study," *Circ. Res.*, vol. 64, no. 3, pp. 449–462, 1989.
- [6] P.-S. Chen, K. M. Moser, W. P. Dembitsky, W. R. Auger, P. O. Daily, C. M. Calisi, S. W. Jamieson, and G. K. Feld, "Epicardial activation and repolarization patterns in patients with right ventricular hypertrophy," *Circulation*, vol. 83, pp. 104–118, 1991.
- [7] A. M. Yue, J. R. Paisey, S. Robinson, T. R. Betts, P. R. Roberts, and J. M. Morgan, "Determination of human ventricular repolarization by noncontact mapping; validation with monophasic action potential recordings," *Circulation*, vol. 110, pp. 1343–1350, 2004.
- [8] M. Potse, B. Dubé, J. Richer, A. Vinet, and R. M. Gulrajani, "A comparison of monodomain and bidomain reaction-diffusion models for action potential propagation in the human heart," *IEEE Trans. Biomed. Eng.*, vol. 53, no. 12, pp. 2425–2435, 2006.
- [9] D. Durrer, R. T. van Dam, G. E. Freud, M. J. Janse, F. L. Meijler, and R. C. Arzbaecher, "Total excitation of the isolated human heart," *Circulation*, vol. 41, no. 6, pp. 899–912, 1970.
- [10] K. H. W. J. ten Tusscher, D. Noble, P. J. Noble, and A. V. Panfilov, "A model for human ventricular tissue," *Am. J. Physiol. Heart Circ. Physiol.*, vol. 286, pp. H1573–H1589, 2004.
- [11] P. G. A. Volders, K. R. Sipido, E. Carmeliet, R. L. H. M. G. Spätsjens, H. J. J. Wellens, and M. A. Vos, "Repolarizing  $K^+$  currents  $I_{TO1}$  and  $I_{Ks}$  are larger in right than left canine ventricular midmyocardium," *Circulation*, vol. 99, pp. 206–210, 1999.
- [12] J. Di Diego, J. Cordeiro, R. J. Goodrow, J. M. Fish, A. C. Zygmunt, G. Pérez, F. Scornik, and C. Antzelevitch, "Tonic and cellular basis for the predominance of the Brugada syndrome phenotype in males," *Circulation*, vol. 106, pp. 2004–2011, 2002.
- [13] B. J. Roth, "Electrical conductivity values used with the bidomain model of cardiac tissue," *IEEE Trans. Biomed. Eng.*, vol. 44, pp. 326–328, 1997.